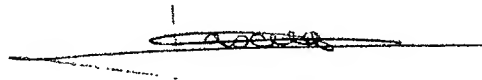


**VERIFICATION OF TRANSLATIONS**

Lise PASCUAL, residing at 6 bis Chemin du Rouquier, 13800 Istres, France,  
declares :

- (1) that she knows well both the French and English languages,
- (2) that she translated the priority documents FR03/08289 and FR02/13022 of the above-identified Application from French to English,
- (3) that the attached English translations are true and correct translations of the priority documents FR03/08289 and FR02/13022 to the best knowledge and belief, and
- (4) that all statements made of her own knowledge are true and correct statements made of information and belief are believed to be true, and further that these statements are made with the knowledge that willful false statements and the like are punishable by fine or imprisonment, or both, under 18 U.S.C § 1001, and that such false statements may jeopardize the validity of the application or ant patent issuing thereon.

Date: September 10, 2010



## TRANSLATION OF PRIORITY FR 0213022

### Screening of molecules with anti-prion activity: kits, methods and screened molecules

5

#### Abstract

The present invention concerns screening molecules with anti-prion activity. More particularly, it concerns kits for screening molecules with anti-prion activity characterized in that they comprise in combination a [PSI+] phenotype yeast, an antibiogram, the screening methods, and a family of molecules with anti-prion  
10 activity isolated by the inventive screen. The invention is applicable to anti-prion agents for producing medicines in particular for treating neurodegenerative diseases involving protein aggregates.

#### Description

15 The present invention relates to screening of molecules with anti-prion activity. It relates more particularly to kits for screening molecules with anti-prion activity, methods of screening, and a family of molecules with anti-prion activity revealed using the screen according to the invention.

20 Prions are infectious proteins responsible for certain neuro-degenerative diseases of spongiform encephalopathy type in mammals, such as Creutzfeldt-Jakob's disease in humans or also the so-called "mad cow disease" in bovines or "scrapie" in ovines. These different diseases are caused by unconventional infectious agents: unlike traditional infectious agents (bacteria, viruses for  
25 example), they contain no nucleic acids. Professor Stanley Prusiner formulated the "protein-only" hypothesis, according to which the infectious agent would be constituted only by a protein. This protein exists naturally in cells in a "normal" (or PrP<sup>c</sup>) form, i.e. soluble, essentially in the form of an  $\alpha$  helix and non-aggregated, therefore functional. Under certain still unknown conditions, this  
30 protein can be converted to a prion (or PrP<sup>sc</sup>) form. In this prion form, the

protein forms insoluble aggregates, essentially in the form of  $\beta$  sheets. The infectious character of this PrP<sup>sc</sup> prion conformation would result from the fact that, apart from the characteristics indicated previously, the protein in prion form also gains the ability to catalyze the passage from the normal Prp<sup>c</sup> cell form to the PrP<sup>sc</sup> prion form in a "snowball"-type mechanism.

Baker's yeast *Saccharomyces cerevisiae* contains several proteins that behave like prions (Fernandez-Bellot and Cullin, 2001). Since as long ago as the 1960s, two unconventional genetic mechanisms have been described. In 1994, the corresponding [PSI+] and [URE3] phenotypes were proposed as resulting from the autocatalytic inactivation of the Sup35p and Ure2p proteins respectively. These prion proteins therefore have a priori a mechanistic analogy with mammal systems deleterious to public health. Like the PrP protein, the "normal" Sup35p protein passes from a soluble state to an insoluble and aggregated state as soon as the protein is in contact with another Sup35p protein in prion form. This aggregated state is verified both by centrifugation experiments and by intracellular localization experiments. Yeast prions can be eliminated ("cured") by a strong dose (3 to 5 mM) of guanidium chloride. As a result of such a treatment (which must applied to at least six to ten generations), the protein aggregates generated by the presence of the prions disappear and the protein in question (Sup35p, for example) is found in a normal, soluble, functional form but having retained the capability of being converted to a prion form should it again come into contact with another Sup35p protein in such a state.

25

The Sup35p protein, in a heterodimeric complex with the Sup45p protein, forms a translation termination factor. This factor recognizes the opal stop codons (UGA). In its normal cell form (soluble and active) in the [psi-] strains, Sup35p, in combination with Sup45p effectively terminates translation at the level of these opal codons. In a [PSI+] strain where the Sup35p protein is in prion form, it is mostly present in the form of insoluble aggregates. Being unable to bind to Sup45p, it is thus non-functional in the translation termination. A small fraction of all of the cellular Sup35p proteins however remains soluble in these [PSI+]

30

cells where it makes it possible, in a complex with Sup45p, to ensure a "minimum translation termination service", a service essential to the survival of the yeast. A colorimetric system making it possible to detect, in an indirect fashion, the form in which the Sup35p protein is present: normal or prion, has  
5 been produced from these findings. This system, which has been described for a long time (see the article on synthesis by Fernandez-Bellot and Cullin, 2001), is based on the use of the *adel-14* allele of the *ADE1* gene, coding for an enzyme of the adenine biosynthesis route: SAICAR synthetase. This enzyme catalyzes the formation of 4-(N-succinocarboxamide)-5-aminoimidazole  
10 ribonucleotide (SAICAR) from 4-carboxy-5-aminoimidazole ribonucleotide (CAIR). The *adel-14* allele contains an opal codon in the reading frame of the *ADE1* gene. In a [psi-] strain, Sup35p in combination with Sup45p will therefore stop the translation of the *ADE1* gene at the level of this stop codon. The protein *adel-14p* thus translated will be truncated and therefore non-functional.  
15 As a result the substrates upstream of the Ade1p enzyme will accumulate, in particular the 5-aminoimidazole ribonucleotide (AIR) The AIR being oxidized to a red-coloured compound, the colonies formed by the [psi-] cells will be red in colour. Moreover, these cells will be auxotrophic for adenine. Conversely, in a [PSI+] strain, the protein Sup35p is essentially present in the form of  
20 aggregates therefore incapable of being combined with Sup45p in order to stop translation at the level of the opal codon of the *adel-14* allele of the *ADE1* gene. As a result, the ribosomes will pause at the level of this stop codon before resuming their translation activity (readthrough). A certain quantity of functional Ade1p protein will therefore be synthesized, the cells will be autotrophic for  
25 adenine and will form white to pink-coloured colonies.

In an article which appeared in P.N.A.S, Prof. Stanley Prusiner's team discloses a test for detecting molecules with anti-prion activity (Korth et al., 2001). This test is carried out on a mammal model (murine neuroblastomas infected with  
30 PrPsc). The safety conditions (P3 laboratory) and cell culture conditions (significant handling) do not allow high-throughput screening to be carried out.

The Application WO 98/30909 also describes a process for screening

molecules with anti-prion activity carried out on rodents infected with an unconventional transmissible agent. This screening method has the same limits as the method described in P.N.A.S.

- 5 The inventors' work has led them to produce a high-throughput screening system in order to detect molecules possessing an anti-prion activity, based on the calorimetric reporter system of the protein Sup35p, described above.

The present invention therefore relates to a kit for screening molecules with an  
10 anti-prion activity, characterized in that it comprises in combination a yeast of phenotype [PSI+] and an antibiogram.

Although based on yeast prions, the kit according to the invention makes it possible to isolate molecules active against mammal prions. Example 7 below  
15 shows that the most active molecules isolated by Prof. Prusiner also have an activity in the screen according to the invention.

However, numerous differences are observed between yeast prions and mammal prions. In an article in the journal "Cellular and Molecular Life  
20 Sciences", Professor C. Cullin proposes, even in view of these differences, distinguishing yeast prions from mammal prions by using the term "propagons". As particular differences between "prions" (mammal) and "propagons" (yeast), there can be mentioned the cytoplasmic character of propagons whereas the mammal PrP prion is a protein anchored to the plasmic membrane, the  
25 pathological character of mammal prions, as well as a certain number of biophysical differences (ternary and quaternary structure, reversibility of the curing etc.)

One of the main advantages of such a screen resides in its complete  
30 harmlessness which allows it to be carried out in a standard level L2 molecular biology laboratory, and not, as required in the previous techniques, in a level P3 laboratory.

Moreover, the great ease of use and very low cost of this kit make it possible carry out high-throughput screening. The use of antibiogram pellets, which allow the diffusion of the product by creating a concentration gradient, moreover makes it possible to test a multiplicity of concentrations in a single experiment, unlike the standard tests, in which only one concentration is tested. For each molecule the anti-prion activity of which is tested, the use of the antibiogram also makes it possible to acquire information on the toxicity of the product as well as on the activity/concentration ratio, and thus to determine the best effective concentration.

10

According to a preferred embodiment, the *[PSI+]* strain used in the kit according to the invention carries an inactivation of the *ERG6* gene.

In fact, yeasts are naturally fairly impermeable. In particular, the preferred yeast for implementing the invention, *Saccharomyces cerevisiae*, has an impermeability such that the carrying out of a screening process proves particularly ineffective without this inactivation.

The screen analysis method according to the invention is visual. According to the anti-prion activity of the molecule tested, the colonies of cells will have a red, pink or white staining. The choice of the strain of yeast can make it possible to improve the contrast between the colonies. In fact, certain so-called "Strong" strains facilitate visual analysis of the screen. Such strains possess a strong level of aggregation of the prion forms. In the opposite case, the strain is referred to as "Weak". The strains preferred for implementation of the invention are therefore the "Strong"-type strains.

Other yeasts can also be used. As examples there can be mentioned: *Kluyveromyces lactis*, *Pichia methanolica*, *Saccharomyces ludwigii*, *Kluyveromyces marxianus*, *Pichia pastoris*, *Zygosaccharomyces rouxi*, *Schizosaccharomyces pombe*.

Given the synthetic lethality observed between the inactivation of the *ERG6*

gene and the inactivation of the *TRP1* gene, the *ERG6* gene can be deleted using the *TRP1* gene as deletion marker.

Advantageously, the kit moreover comprises a prion curing agent at sub-effective  
5 effective doses.

By curing, is meant an elimination of the prion forms from the yeast cells. This elimination can be temporary or permanent.

10 By way of example, a prion curing agent can be hydrogen peroxide or preferentially, guanidium chloride.

By sub-effective doses, is meant doses which used alone would not suffice to eliminate the prions from the yeasts. The values of such doses are given, in the  
15 examples which follow, for guanidium chloride.

The benefits of the presence of a curing agent at sub-effective doses are to reinforce the sensitivity of the screen and obtain a better contrast.

20 The kit according to the invention can be used in a method for screening molecules with anti-prion activity. This screening method, to which the invention also relates, is characterized in that it uses a [PSI<sup>+</sup>] phenotype yeast and comprises the following stages:

- a. production of a lawn of cells in vitro with a sub-effective dose of a prion  
25 curing agent,
- b. deposition of the compounds to be tested according to the antibiogram method,
- c. incubation for approximately 2-4 days at approximately 20-25°C, and,
- d. analysis of the staining of the cell colonies.

30

This method possesses advantages analogous to those of the kit according to the invention. It is a visual test, very easy to analyze. Its implementation is very simple and inexpensive. The precautions relative to safety are those of a

standard molecular biology laboratory. It allows mass screening: a single person can manually screen more than 400 products per day. Very high-throughput screening would be possible by automation of the method. The screen result is developed after 7 days, without it being necessary to resort to a lot of handling between day D and day D+7 (optionally a change in temperature of the incubator). Finally, this method is particularly economical.

According to a preferred embodiment, the screening method, according to the invention, is characterized in that yeast *ERG6* gene is inactivated. One of the yeasts preferred for the implementation of this method is *Saccharomyces cerevisiae*.

Advantageously, the curing agent of Stage a. is guanidium chloride.

The method can also comprise the following stages:

- e. incubation for approximately 2-4 days at approximately 2-6° C., and/or,
- f. carrying out a secondary screening test.

The incubation at 2-6° C. makes it possible to accentuate the contrast in staining of the colonies.

Preferentially, the secondary screening test can comprise the following stages:

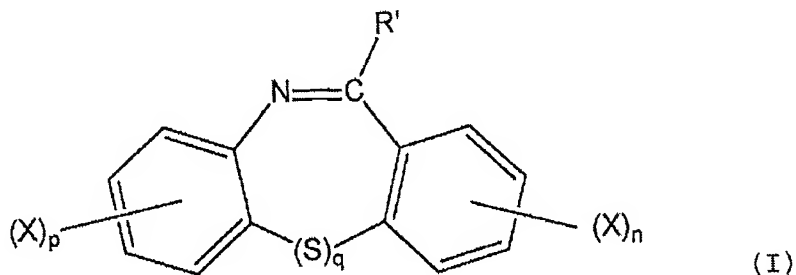
- construction of a strain of yeast in which the ADE2 gene is under the control of the DAL5 gene promoter
- carrying out Stages a. to e. of the methods described above, the stage a. comprising moreover one sub-effective dose of guanidium chloride.

Such a secondary screening makes it possible to test very rapidly whether the molecules isolated during the primary screening can have a general effect on the prions in the yeast. In fact, the SUP35 genes (responsible for the [PSI<sup>+</sup>] prion) and URE2 (responsible for the [URE3] prion) code for enzymes having totally different functions and the primary sequences of which are very remote.



The invention also covers the molecules isolated by the screening method according to the invention.

In particular, the screening method has made it possible to isolate anti-prion agents having the following formula (I):



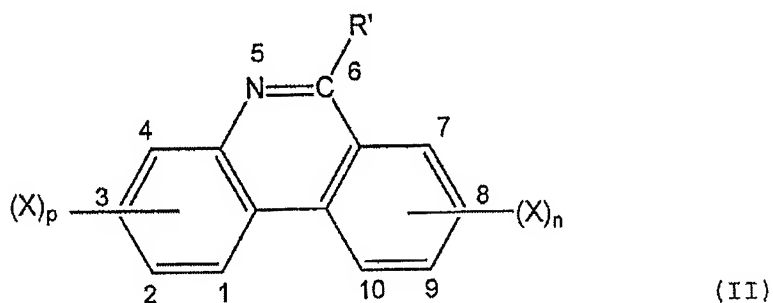
in which R' is an H, NH<sub>2</sub>, NHR<sup>2</sup> group, where R<sup>2</sup> is an alkyl or alkylaminoalkyl chain with 1 to 10 carbon atoms, branched or unbranched,

X represents F, Cl, Br, I, CF<sub>3</sub>, SR<sup>3</sup>, OR<sup>3</sup>, OH, NO<sub>2</sub>, COR<sup>3</sup>, CONH<sub>2</sub>, COOH, COOR<sup>3</sup>, where R<sup>3</sup> is an alkyl group with 1 to 4 carbon atoms, preferably CH<sub>3</sub>.

p and n, identical or different, are equal to 0, 1 or 2, q is equal to 0 or 1,

q is equal to 0 or 1.

The invention relates in particular to the anti-prion agents of formula (II):



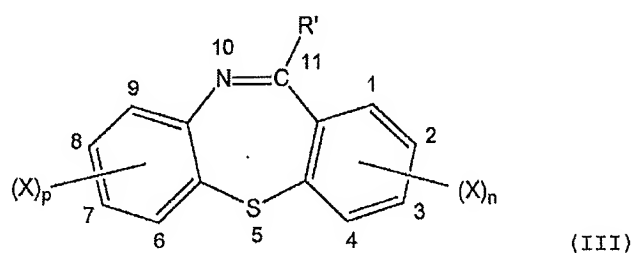
in which R' represents an H, NH<sub>2</sub>, NH-(CH<sub>2</sub>)<sub>3</sub>-N(CH<sub>3</sub>)<sub>2</sub>, NH-CH(CH<sub>3</sub>)-(CH<sub>2</sub>)<sub>3</sub>-N(CH<sub>2</sub>-CH<sub>3</sub>)<sub>2</sub> group,

X represents F, Cl,

p and n, identical or different, are equal to 0, 1 or 2.

Some compounds of this family are particularly active. It is phenanthridin and 6-aminophenanthridin, and their chlorinated derivatives, in particular when the chlorine is placed in position 8, 9, 10, and preferentially in position 10 (see in the following examples).

The invention relates more particularly to the compounds of formula (III):



in which R' represents an H, NH<sub>2</sub>, NH-(CH<sub>2</sub>)<sub>3</sub>-N(CH<sub>3</sub>)<sub>2</sub>, NH-CH(CH<sub>3</sub>)-(CH<sub>2</sub>)<sub>3</sub>-N(CH<sub>2</sub>-CH<sub>3</sub>)<sub>2</sub> group,

X represents F, Cl,

p and n, identical or different, are equal to 0, 1 or 2.

This family of molecules, called "Kastellpaolitines" by the inventors, possesses the sought anti-prion activity to a greater or lesser degree. In particular, the chlorinated derivatives of this family are particularly effective. The best effectivenesses are obtained when chlorine is placed in position 2, 3 or 4, preferably in position 4 (see KP1 in the examples which follow).

20

The anti-prion agents according to the invention are particularly useful for obtaining a medicament intended to prevent and/or to treat neurodegenerative diseases, in particular of the protein-aggregation type, such as the spongiform encephalopathies, Alzheimer's and Huntington's disease etc. These medicaments can be intended for human or veterinary use, in particular for domestic (cows, sheep etc.) or wild animals (lynx, the Cervidae such as deer, moose etc.).

25

Other characteristics and advantages of the invention will become apparent in the examples below and by referring to the following figures:

- FIG.1 relates to the feasibility of the screen,
- FIG.2 illustrates the screening protocol,
- 5       - FIG.3 relates to the isolation of the Kastellpaolitines, phenanthridine and to their structure/activity relationship,
- FIG. 4 relates to the determination of the 6-aminophenanthridin activity,
- FIG. 5 shows the results of the liquid curing tests,
- FIG. 6 relates to the secondary screen based on the [URE3] prion,
- 10       - FIG. 7 demonstrates the validation of the test with chlorpromazine, quinacrine.

#### **EXAMPLE           1:           Carrying           Out           the           Screen.**

##### **15   1. Material and Methods Organisms (*Saccharomyces cerevisiae*) and Media Culture**

The [PSI+] haploid yeast strain 74-D694 (Mat a, adel-14, trpl-289, his3-Δ200, ura3-52, leu2-3,112) was used in the development of the screening method.

20   The strain used is called "Strong" as it has a well-marked phenotype when the translation termination factor Sup35p is in prion or aggregated form.

In order to increase the penetration of the inhibitors, the inventors genetically modified this strain by introducing into it a mutation of the ERG6 gene. This

25   gene is involved in the biosynthesis of ergosterol, a component of the cell wall of the yeasts. The mutation was produced by insertion at the level of the chromosome site of the ERG6 gene of a "deletion cassette" corresponding to the TRP1 marker gene flanked by DNA sequences situated upstream and downstream of the coding frame of the ERG6 gene. This cassette was

30   produced by PCR using the plasmid pFA6a-kanMX6 as matrix and the oligonucleotides oBM1060 (5') et oBM1061 (3') as primers. The "Strong" 74-D694 yeast cells having integrated the deletion cassette (strain called STRg6, deposited at the CNCM on 10th Oct. 2002 under number 1-2943) are those

which develop on minimum media devoid of tryptophan. The mutation  $\Delta$ erg6: TRP1 was then verified by PCR using the genomic DNA of the strain STRg6 as matrix and the oligonucleotides oBM1030 (5') and oBM1063 (3') as primers.

- 5 The PCR primers used have the following nucleotide sequences: oBM1060 5' CGATTTAAGTTTTACATAATTTAAAAAACAAGAATAAAATAATAATATAGTA GGCAGCATAAGCGGATCCCCGGGTAAATTAA 3' (SEQ ID No. 1)  
oBM1061 5' CTGCATATATAGGAAAATAGGTATATATCGTGCGCTTTATTTGAATCTTATT  
10 GATCTAGTGAATGAATTCGAGCTCGTTTAAAC 3' (SEQ ID No. 2)  
oBM1030 5' GGTACCTCGTTCCCGTAC 3' (SEQ ID No. 3)  
oBM1063 5' CAGTCAGAAATCGAGTTCCA 3' (SEQ ID No. 4)

- Unless otherwise indicated, the yeast strains are cultured at 30°C in rich  
15 medium (YPD $\Psi$ ) or in minimum medium. Unless explicitly specified, the percentages correspond to a mass/volume ratio. The gelosed form is obtained by the addition of 2% agar.

- YPD $\Psi$ :** 1% yeast extract (FISHER®), 2% peptone A (GIBCO®) and 2%  
20 glucose;

- Minimum medium:** 0.175% yeast nitrogen base without amino acid and ammonium sulphate (DIFCO®), 0.75% ammonium sulphate and 2% glucose. This medium is adjusted to pH 6. In order to compensate for possible auxotrophies, this medium can be completed, after sterilization, by the addition  
25 of amino acids (0.002% L-histidine and/or 0.004% L-leucine and/or 0.003% L-tryptophan) or nitrogenous bases (0.0025% uracil and/or 0.008% adenine).

#### **Method for screening substances with anti-prion activity ("Prion Halo Assay")**

- 30 The screening method developed is based on the antibiogram principle. In fact, the compounds to be tested are applied to a sterile filter-paper disc, itself applied to a dish of solid YPD $\Psi$  medium containing 0.2 mM of guanidium chloride previously seeded with approximately  $10^6$  cells of the STRg6 strain in

order to produce a yeast lawn. The addition of a small quantity of guanidium chloride (0.2 mM), a sub-effective dose for eliminating prions from yeast (the effective dose being of the order of 3 to 5 mM) makes it possible to increase the sensitivity of the test (see Results section). The 12 cm square dishes are then incubated for 3 days at 23.5°C in order to allow the appearance and growth of the yeast colonies. These dishes are then stored for 3 days at 4°C in order to accentuate the red staining present around the discs soaked with ingredients active on the prion form of the protein Sup35p. Comparison with the negative controls (application of the solvent of the inhibitors tested) and positive controls (application of a 300 mM guanidium chloride solution, causing effective elimination of the Sup35p proteins in prion form) makes it possible to judge the effectiveness of a compound. FIG. 2 illustrates the protocol of the screening method: **(1)** Culture of the STRg6 strain; **(2)** Application and plating with sterile glass beads 3 & 4 mm in diameter, of approximately  $10^6$  cells in exponential growth phase on a dish of solid YPD $\Psi$  medium containing 0.2 mM of guanidium chloride: constitution of the cell "lawn"; **(3)** Application of the sterile filter-paper discs according to a grid allowing the analysis of 32 compounds (including controls) and deposit of 20  $\mu$ l maximum of each of the products to be tested; **(4)** Incubation; **(5)** Scanning of the result obtained; **(6)** Example showing the isolation of a compound having a strong anti-prion activity.

### **Synthesis of 11-aminodibenzo[b,f][1,4]thiazepines and 6-aminophenanthridine**

11-aminodibenzo[b,f][1,4]thiazepines, also called Kastellpaolitines, can be prepared in a single stage. The synthesis of these products has already been described in the publication by Mettey and al., 1997.

## **2. Results**

**Principle and Feasibility of the Screen**  
Guanidium chloride, the only product known to effectively eliminate prions from the yeast *Saccharomyces cerevisiae*, served not only as a positive control throughout screening, but also for studying the feasibility of the method as well

as developing it. Guanidium chloride effectively eliminates the different yeast prions at a dose comprised between 3 and 5 mM (Fernandez-Bellot and Cullin, 2001). Under these conditions, the curing requires a constant presence of this product for six to ten generations in exponential growth phase compromising  
5 the feasibility of the screen on a dish such as the inventors wished to achieve.

FIG. 1 shows the feasibility of the screen. The three left-hand panels: a [PSI<sup>+</sup>] strain is cultured for 48 hours in the presence of 5 mM guanidium chloride (with 0.2% DMSO final) or, as a control,  
10 with only 0.2% DMSO final. At T=0, then every 24 hours, a 10 µl drop (approximately 10<sup>4</sup> cells) is applied to a dish of rich medium. The guanidium chloride curing begins to have an effect after 24 hours of treatment, i.e. after approximately 6 generations (a pink staining begins to appear). After 48 hours, i.e. after approximately 12 generations, the drop of cells has a clearly red  
15 staining, a sign of a complete curing of the [PSI<sup>+</sup>] cells. The middle panel: a few cells are taken at T=48 hours and scratched onto a fresh medium. Almost all of them form red colonies in the case of curing with guanidium chloride. The right-hand panel: these same cells are pelleted at the bottom of an  
20 Eppendorf tube after liquid culture. In the case of curing with guanidium chloride, they form a red pellet.

The first stage therefore consisted of determining whether guanidium chloride could have an effect which can be visualized on a dish of [PSI<sup>+</sup>] cells with the  
25 antibiogram pellet system. Once this stage was carried out, the inventors developed the optimum temperature, medium and density conditions as well as cell type to use (FIG. 2). The strain having the best sensitivity is the STRg6 strain cultured at 23.5°C and in the presence of 200µM of guanidium chloride. In fact, the introduction of a sub-effective dose of guanidium chloride into the  
30 medium makes it possible to increase the sensitivity of the test.

### **Screening of a Combinatorial Library**

Compounds (approximately 1000) were passed through the screen using the conditions optimized by the inventors (FIG. 2). On each dish, 15 µl of DMSO are deposited on the filter at the top left (negative control) and 15 µl of a 300 mM solution of guanidium chloride in DMSO (positive control) were applied to the filter at the bottom right.

The same volume (15 µl) of each of the products of the library (all in 10 mM solution in DMSO) was applied to the remaining filters (thirty for each large square Petri dish). A positive signal (visualization of a red halo around the sterile filter-paper disc to which the product is applied) was obtained for five products. These products correspond to four molecules of the same family, called "Kastellpaolitines" by the inventors, and to a well-known fifth molecule: phenanthridine.

## **EXAMPLE 2 : Identification of the Kastellpaolitines and Phenanthridine**

The chemical structures of the Kastellpaolitines and phenanthridine are shown in FIG. 3B. The panel 3A shows a comparative analysis of the size of the red halos obtained with all of these molecules respectively (all applied in an equivalent quantity: 15 µl of a 10 mM solution in DMSO). This experiment makes it possible to compare the relative activity of each of these products. The most active is Kastellpaolitine 1 (or KP1) followed by phenanthridine.

	<b>6-aminophenanthridine</b>	<b>Synthesis</b>	<b>and</b>	<b>Test</b>
25	Comparative analysis of phenanthridine on the one hand, and of the Kastellpaolitines on the other hand show several common points between these two groups of molecules (FIG. 3). The different molecules are classified there from the least active to the most active and their respective formulae indicated. All are tri-cyclic, the central ring containing in all cases a nitrogen atom with a double bond to an adjacent carbon atom. In contrast, in all the Kastellpaolitines, the carbon of the central ring which has a double bond to this nitrogen atom carries an amino group, which is not the case for phenanthridine. This observation led the inventors to want to test 6-aminophenanthridine.			

6-aminophenanthridine can be prepared according to the procedure developed by Kessar et al., 1969. 6-aminophenanthridine was therefore passed through the screen according to the invention, in comparison with the Kastellpaolitines 1 (KP1) and 5 (KP5) as well as phenanthridine. The result is very clear: 6-aminophenanthridine is still more active than the Kastellpaolitines and phenanthridine.

FIG. 4 illustrates the results of this comparison: the activity of 6-aminophenanthridine was determined on a dish and compared to KP1 and 5 and phenanthridine. For all the molecules, the same quantity is applied (10  $\mu$ l of a 10 mM solution). In the case of the positive control (guanidium chloride), the solution used was 300 mM.

As a result, by grafting this amino group, characteristic of the Kastellpaolitines onto phenanthridine, the inventors significantly increased the activity of the latter.

### **EXAMPLE 3 : Synergy Between Products Isolated Using the Screen and Guanidium Chloride**

All the active molecules were isolated in a medium containing a weak dose of guanidium chloride (200 $\mu$ M/effective dose=4 mM). Taking this course, established during the development of the screen corresponded to the wish to increase the sensitivity (and therefore the detection threshold of the method). The effect of the molecules in media containing more (500  $\mu$ M) guanidium chloride or not containing any, was observed subsequently. Phenanthridine is always active on a medium without guanidium chloride, but its activity increases significantly as a function of the quantity of guanidium chloride (however in a clearly sub-effective dose) in the medium. This result indicates a synergy of action between guanidium chloride and phenanthridine. The same result was obtained for all the other molecules isolated by the inventors (the Kastellpaolitines and 6-aminophenanthridine).



#### **EXAMPLE 4 Verification of Liquid Medium Curing**

The inventors then wanted to determine whether the red halos observed in the yeast test corresponded to [PSI<sup>+</sup>] prion curing and not to an artefact (for example these red halos could be due to a direct inhibition of the biosynthesis chain of adenine by these molecules, which would lead to a accumulation of the AIR). If these molecules effectively eliminate the [PSI<sup>+</sup>] prion, a treatment of [PSI<sup>+</sup>] cells in liquid culture followed by washing of said cells must allow them to form red colonies on a gelosed medium no longer containing the molecules. These tests were carried out with 6-aminophenanthridine on the wild-type "strong" strain 74-D694.

The liquid medium curing conditions are the following: a [PSI<sup>+</sup>] strain is cultured for 5 days in liquid medium in the presence of the indicated quantities of the different products (see FIG. 5). Every 24 hours, an aliquot fraction is washed in medium uncontaminated by any product and applied to a solid gelosed medium (itself also uncontaminated by any product) which is then treated as indicated in FIG. 2.

As shown in FIG. 5, 6-aminophenanthridine is capable of partially curing the [PSI<sup>+</sup>] prion from a significant number of cells. The curing effectiveness can in particular be increased by adding a sub-effective dose (100  $\mu$ M) of guanidium chloride to the culture medium. In such a liquid curing, the same synergic effect as that observed in the dish test is also found.

25

#### **EXAMPLE 5 Development and Use of a Secondary Calorimetric Screen Based on the Use of [URE3], Another Yeast Prion.**

Another rapid dish test was carried out, based on another yeast prion: [URE3]. This test constituted a secondary screen which makes it possible to generalize the effect of the products isolated during the primary screen of another yeast prion. In this way, it is possible to remove the molecules active only against the [PSI<sup>+</sup>] prion and therefore less useful, having a non-general effect.

30

For the [URE3] prion the haploid strain used is CC34 (*Mat  $\alpha$ , trp1-1, ade2-1, leu2-3,112, his3-11, 15, ura2:: HIS3*).

- 5 The **NT34** strain which served for the secondary screen was constructed from CC34, a strain in which the coding frame of the *DAL5* gene has been replaced by that of the *ADE2* gene using the same method as that used for the construction of the **STRg6** strain. For this purpose a deletion cassette corresponding to the *ADE2* gene flanked by DNA sequences situated upstream and downstream of the coding frame of the *DAL5* gene was produced by PCR  
10 using genomic DNA of the BY4742 haploid strain (*Mat  $\alpha$ , his3 $\Delta$ 1, leu2  $\Delta$ O, lys2  $\Delta$ O, ura3 $\Delta$ O*) as matrix and the oligonucleotides: TATATTCTTCTCTGATAACAATAATGTCAGTGTATCTCACCACTATTATTACT TGTT TTCTAGATAAGC (SEQ ID No. 6) (3') as primers.
- 15 The mutation *DAL5::ADE2* was then verified by PCR using the genomic DNA of the NT34 strain as matrix and the oligonucleotides: ATAGTCTCTGCTCATAG (SEQ ID No. 7) (5'), and GCTTACAGAAATTCTAC (SEQ ID No. 8) (3') as primers. The NT34 strain (*Mat  $\alpha$ , trp1-1, ade2-1, leu2-3,112, his3-11,15, ura2::HIS3, DAL5::ADE2*) was deposited at the CNCM on 10th Oct. 2002 under number 1-2942.
- 20

- This screen is based on the same colorimetric system as the primary screen. In the **NT34** yeast strain, the *ADE2* gene is no longer under the control of its own  
25 promoter, but under that of the *DAL5* gene. When the protein Ure2p is in prion form ([URE3]), the transcription from the promoter of the *DAL5* gene is activated, therefore the *ADE2* gene is expressed, therefore the strains are white and autotrophic for adenine. When the URE2p protein is in the normal form ([URE3-0]), the transcription from the promoter of the *DAL5* gene is repressed,  
30 therefore the *ADE2* gene is not expressed, therefore the strains are red and auxotrophic for adenine. When the NT34 strain is treated with 5 mM of guanidium chloride for approximately ten generations, it forms red colonies (as expected and as the [PSI+] strain used for the primary screening would do). As

can be observed in FIG. 6, phenanthridine and 6-aminophenanthridine cause the appearance of a red halo when they are applied to the small filter itself applied to the lawn of cells previously plated on the gelosed nutritive medium (same process as for the primary screen, see FIG. 2). This result suggests that these products are also active on the [URE3] prion. It is to be noted, however, that this secondary screen is clearly less sensitive than the primary screen. It is therefore very useful for rapidly observing whether the effect of the molecules isolated during the first screen can be generalized to other yeast prions but in no event could it be substituted for the primary screen.

#### **EXAMPLE 6 Verification of Liquid Medium [URE3] Curing**

Two types of experiments were carried out in order to verify that the effect observed on dishes with the NT34 strain corresponds to curing. Firstly, cells in the zones surrounding the filter were recovered for the negative (DMSO), and positive (guanidium chloride) control for phenanthridine and for 6-aminophenanthridine. These cells were then scratched onto a fresh medium free of all these molecules. The cells recovered around the filters all form red colonies, with the exception of those collected around the negative control. This result shows that the red staining observed on dishes for the NT34 strain corresponds to curing and not to an artefact linked to inhibition of an enzyme of the biosynthesis route of adenine (in this case, the red staining would be lost on a medium without inhibitor). The curing effect of phenanthridine and 6-aminophenanthridine was also directly verified on the [URE3] prion. [URE3] cells of the CC34 strain grow on a medium called USA whereas cured ([ure3-0]) cells are incapable of growing on this medium.

The inventors examined the ability of [URE3] cells treated with 200  $\mu$ M of guanidium chloride (negative control), 5  $\mu$ M of guanidium chloride (positive control) or with different doses of 6-aminophenanthridine (alone or in combination with 200  $\mu$ M of guanidium chloride) to grow on a USA medium. 6-aminophenanthridine is capable of curing the [URE3] prion in a significant manner and, just as for the [PSI+] prion, this effect is accentuated by a low dose of guanidium chloride (200  $\mu$ M). These results, apart from the fact that they

validate the secondary screen with the NT34 strain, suggest that the effect of the inhibitors revealed by said screen should be general on all yeast prions.

**EXAMPLE 7 Validation of the Screen with Two Molecules Active on the**

5 **Mammal Prion PrP: Chlorpromazine and Guinacrine**

The laboratory of Stanley Prusiner, who first put forward the "protein-only" hypothesis and was awarded the Nobel prize in 1997, has isolated a certain number of molecules active on the mammal prion PrP using a system of murine  
10 cells (neuroblastomas) chronically infected with the prion PrPsc (Korth et al., 2001). This system, due to its labour-intensiveness and its complexity, does not allow mass screening like that developed by the inventors. Thus the approach of Stanley Prusiner's group was to test one-by-one, from the molecules already used as medicaments, those which pass the blood-brain barrier. Certain  
15 molecules, such as in particular quinacrine (used as an anti-malarial drug for a long time) or chlorpromazine (an antidepressant) have a particular activity in their system. In order to validate the screen, the inventors therefore tested chlorpromazine and quinacrine in their yeast system.

As shown in FIG. 7, these two molecules have a certain activity against the  
20 [PSI+] prion. It must however be noted that their activities are clearly weaker than that of 6-aminophenanthridine. It can also be seen that chlorpromazine and quinacrine, like all of the molecules highlighted by the invention, exhibit a strong synergy of action with guanidium chloride (In FIG. 7, the medium used contains 200 µM of guanidium chloride) The latter result suggests that these  
25 two molecules act on the same biochemical route as the isolated molecules according to the invention. Moreover, it is interesting to note that quinacrine, the activity of which is approximately ten times greater than that of chlorpromazine in Prof. Prusiner's test, also exhibits an activity greater than the latter in the screen developed by  
30 the inventors. Moreover, just as in Prof. Prusiner's test, chlorpromazine and quinacrine require prolonged treatment (at least 6 days in the case of Prof. Prusiner's test, at least two to three days in the case of the screen according to the invention) before an activity is detected.

Moreover, the inventors determined the activity, in the test according to the invention, of other molecules isolated using the test based on mouse neuroblastomas, developed by Prof. Prusiner. A good correlation was found  
5 between the results obtained in the two systems: acepromazine which is shown to be slightly active in the mammal system also exhibits a weak activity in the test according to the invention and the molecules inactive in analysis on mammals such as carbamazepine, imipramine, haloperidol, chlorprothixene or methylene blue were also inactive in the test.  
10 All these correlations between the activity of quinacrine and chlorpromazine according to the test or the screen used make it possible to validate the use of the method according to the invention in order to carry out high-throughput screenings with a view to isolating molecules capable of constituting effective medicaments (on mammals and in particular humans) against  
15 neurodegenerative diseases involving protein aggregates, of spongiform-encephalopathy type, Alzheimer's disease, Huntington's disease etc.

### Bibliographic references

**Fernandez-Bellot et al.**, "The protein-only theory and the yeast *Saccharomyces cerevisiae*: the prions and the propagons", CMLS, 5 2001,58:1857-1878.

**Korth C. et al.**, "Acridine and phenothiazine derivatives as pharmacotherapeutics for prion disease", PNAS, 2001, 98(17):9836-9841. 10

**Mettey Y. et al.**, "Synthesis of 11-Aminodibenzo [b, f] [1, 4J thiazepines and Fluoro derivatives" , J. Heterocyclic Chem., 1997,34:465-467.

**Kessar S. V. et al.**, Tetrahedron Letters, 1969,1151.

## Claims

1. Kit for screening molecules with an anti-prion activity, characterized in that it comprises in combination a yeast of phenotype [PSI+] with an antibiogram.  
5
2. Kit according to claim 1, characterized in that the ERG6 gene from the yeast of phenotype [PSI+] is inactivated.
3. Kit according to claim 1 or 2, characterized in that the yeast is  
10 *Saccharomyces cerevisiae*.
4. Kit according to any one of claim 1 to 3, characterized in that said kit comprising moreover a curing agent at sub-effective doses.
- 15 5. Kit according to claim 4, characterized in that the prion curing agent is guanidium chloride.
6. Method for screening molecules with anti-prion activity, characterized in that it implements [PSI+] phenotype yeast and comprises the following stages:  
20
  - a. production of a lawn of cells in vitro on a medium complemented with a sub-effective dose of a prion curing agent,
  - b. deposition of the compounds to be tested according to the antibiogram method,
  - c. incubation for approximately 2-4 days at approximately 20-25° C, and,
  - 25 d. analysis of the staining of the cell colonies.
7. Screening method according to claim 6, characterized in that the ERG6 gene from the yeast is inactivated.
- 30 8. Screening method according to claim 6, characterized in that the yeast is *Saccharomyces cerevisiae*.
9. Screening method according to any one of claims 6 to 8, characterized in that

the stage a. moreover comprises the addition of a sub-effective dose of guanidium chloride.

10. Screening method according to any one of claims 6 to 9, characterized in that it moreover comprises the following stages:

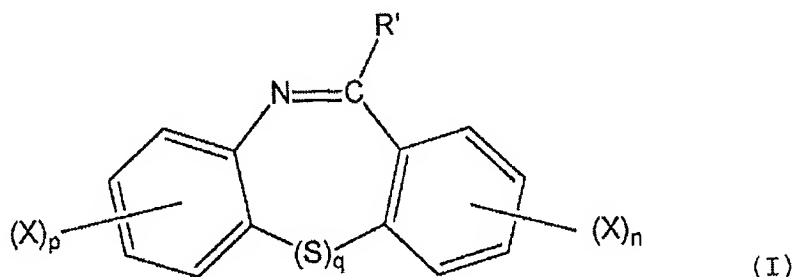
- e. incubation for approximately 2-4 days at approximately 2-6° C., and/or,
- f. carrying out a secondary screening test.

11. Screening method according to claim 10, characterized in that the secondary screening test comprises the following stages:

- construction of a strain of yeast in which the ADE2 gene is under the control of the DAL5 gene promoter
- carrying out Stages a. to e. of the methods according to claims 6 and 10, the stage a. comprising moreover one sub-effective dose of guanidium chloride.

15

12. Anti-prion agent of formula (I):



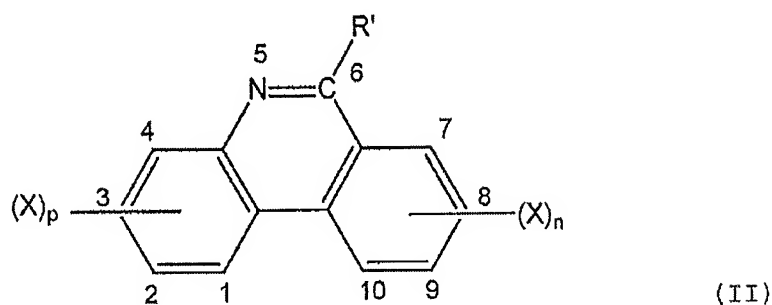
20 in which R' is an H, NH<sub>2</sub>, NHR<sup>2</sup> group, where R<sup>2</sup> is an alkyl or alkylaminoalkyl chain with 1 to 10 carbon atoms, branched or unbranched,

X represents F, Cl, Br, I, CF<sub>3</sub>, SCH<sub>3</sub>, OCH<sub>3</sub>, OH, NO<sub>2</sub>, COCH<sub>3</sub>, CONH<sub>2</sub>, COOH, COOR<sup>3</sup>, where R<sup>3</sup> is an alkyl group with 1 to 4 carbon atoms, preferably CH<sub>3</sub>.

25 p and n, identical or different, are equal to 0, 1 or 2,  
q is equal to 0 or 1.



13. Anti-prion agent according to claim 12, of formula (II) :

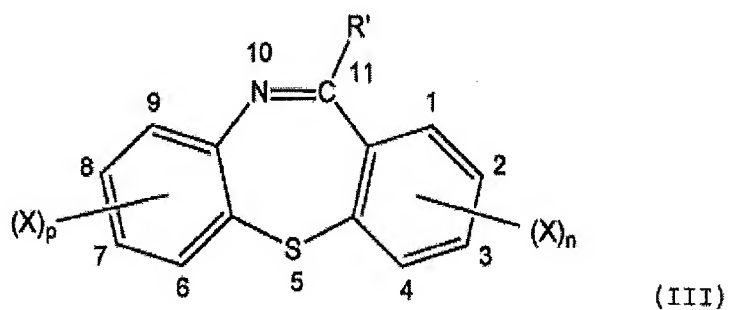


in which R' represents an H, NH<sub>2</sub>, NH-(CH<sub>2</sub>)<sub>3</sub>-N(CH<sub>3</sub>)<sub>2</sub>, NH-CH(CH<sub>3</sub>)-(CH<sub>2</sub>)<sub>3</sub>-N(CH<sub>2</sub>-CH<sub>3</sub>)<sub>2</sub> group,

X represents F, Cl,

p and n, identical or different, are equal to 0, 1 or 2.

14. Anti-prion agent according to claim 13 of formula (III) :



in which R' represents an H, NH<sub>2</sub>, NH-(CH<sub>2</sub>)<sub>3</sub>-N(CH<sub>3</sub>)<sub>2</sub>, NH-CH(CH<sub>3</sub>)-(CH<sub>2</sub>)<sub>3</sub>-N(CH<sub>2</sub>-CH<sub>3</sub>)<sub>2</sub> group,

X represents F, Cl,

p and n, identical or different, are equal to 0, 1 or 2.

15. Use of the anti-prion agents according to any of claims 12 to 14, in order to obtain a medicament intended for treating neurodegenerative diseases involving protein aggregates.

16. Use according to claim 15, characterized in that the diseases are the spongiform encephalopathies, Alzheimer's disease and Huntington's disease.

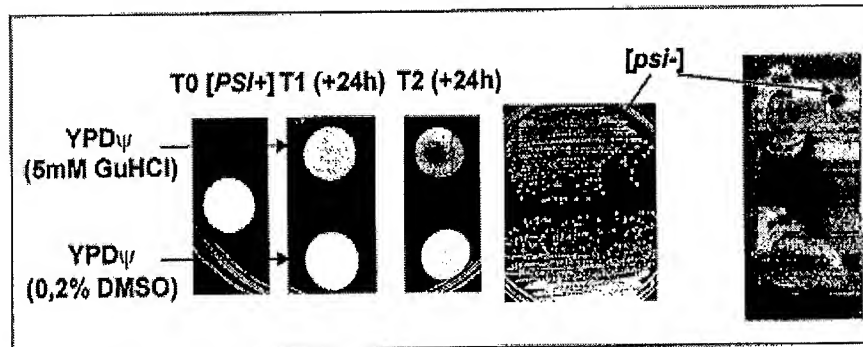


Figure 1

1 night at 30°C

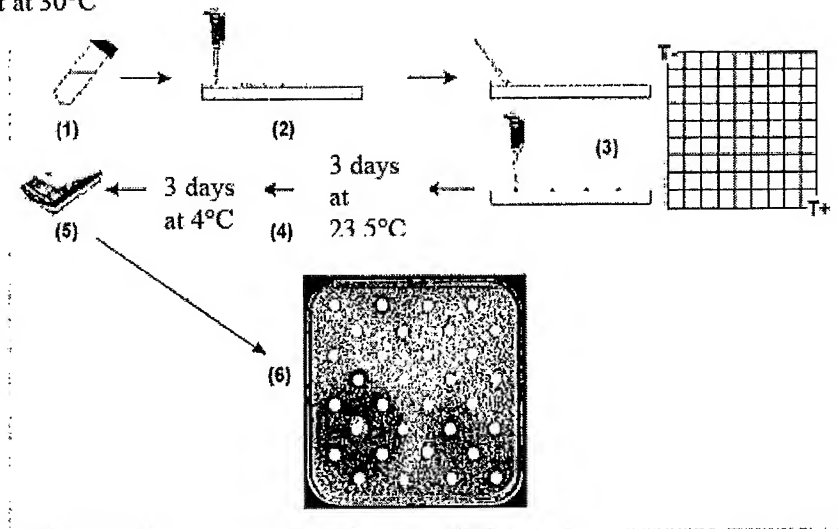


Figure 2

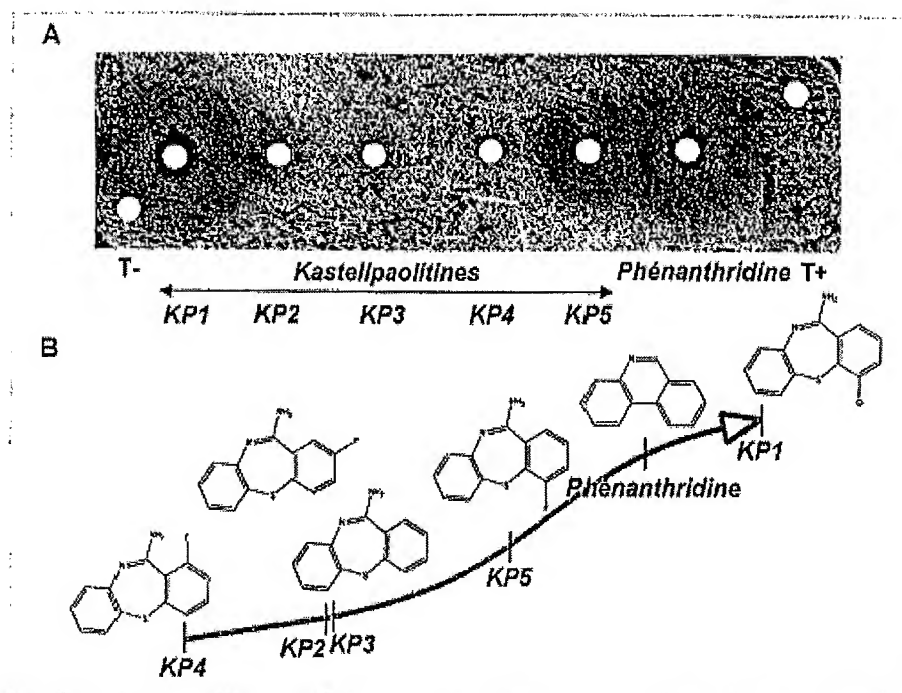


Figure 3

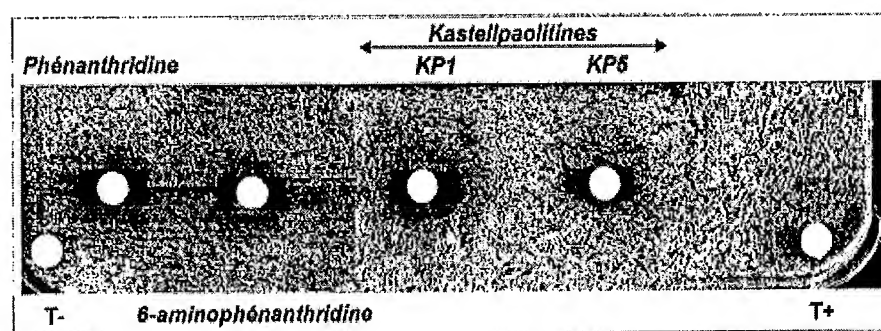


Figure 4

T-  
(100  $\mu$ M GuHCl)

T+  
(4mM GuHCl)

100  $\mu$ M GuHCl  
+ 100  $\mu$ M  
6-aminophénanthridine

100  $\mu$ M GuHCl  
+ 200  $\mu$ M  
6-aminophénanthridine

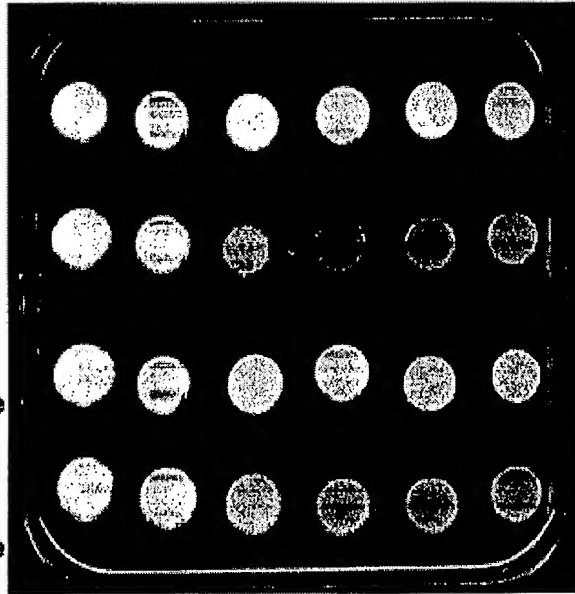


Figure 5

5

10

15

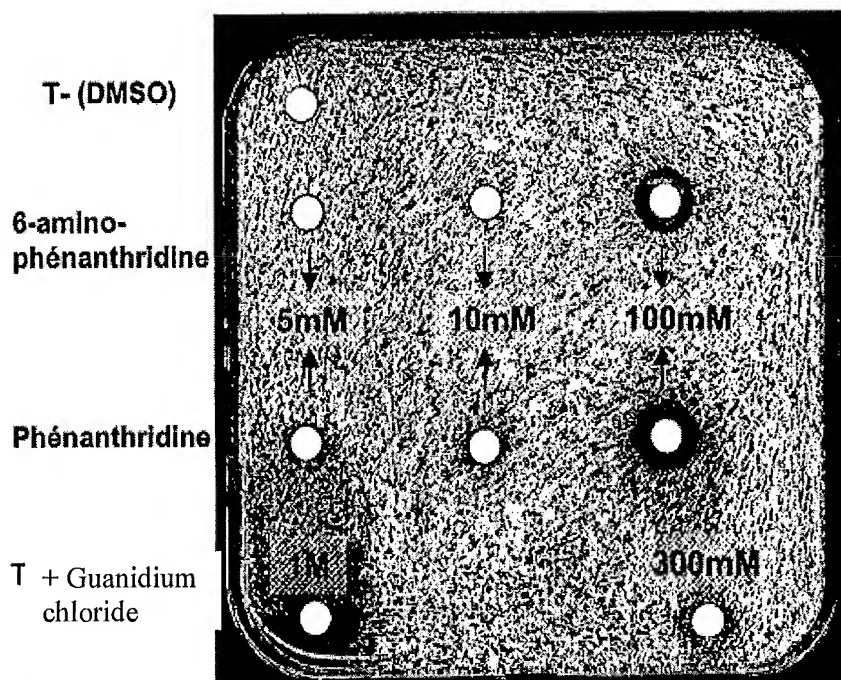


Figure 6

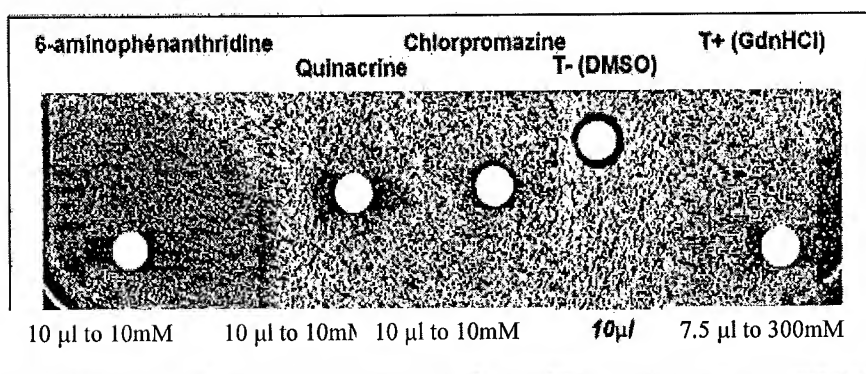


Figure 7

# SEQUENCE LISTING

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kits, methods and screened molecules

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